

Drug Therapy in the Heart Transplant Recipient Part I: Cardiac Rejection and Immunosuppressive Drugs

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Survival after heart transplantation has improved considerably over the past 20 years. Half of all patients now live >9 years, and ≈25% live ≥17 years.¹ Currently, ≈20 000 heart transplant recipients live in the United States.² Improved longevity means prolonged immunosuppression and the concomitant use of drugs to prevent or treat the long-term complications of immunosuppressive agents, such as infection, obesity, hypertension, hyperlipidemia, renal insufficiency, diabetes, osteoporosis, gout, and malignancies. In 1989, heart transplant recipients surviving 1 year were reported to be taking 16±6 drug doses per day (prescription and nonprescription).³ In 2001, heart transplant recipients surviving an average of 76 months were taking 7 prescription drugs (range, 2 to 14), along with a number of nonprescription drugs.⁴ Thus, despite prolonged survival, heart transplant recipients continue to take multiple medications. With the large number of heart transplant recipients in the community and the increasing number of immunosuppressive and non-immunosuppressive drugs used by these patients, it is important that the general cardiologist understand these drugs, their side effects, and the very real potential for drug–drug interactions. These interactions may result in adverse events caused by supratherapeutic and subtherapeutic drug concentrations. In this series, we review mechanisms and types of rejection, immunosuppressive drugs commonly used in the heart transplant recipient, common medical problems after transplantation, and clinically significant drug–drug interactions.

Rejection

A brief review of known immunologic mechanisms leading to graft rejection highlights the action of individual immunosuppressive drugs, as well as the rationale for combination therapy^{5–8} (Figure). The rejection of a transplanted organ is primarily a T-lymphocyte (T-cell)–mediated event, although humoral (B-cell) responses also contribute. The exception is hyperacute rejection, which occurs when preformed antibod-

ies to human leukocyte antigens (HLA) result in an immediate and catastrophic rejection. Immune recognition of donor antigens that differ from those of the recipient (allorecognition) begins with the function of antigen-presenting cells (APCs). APCs are usually dendritic cells, macrophages, or B cells, although other types of cells, particularly endothelial cells, can be stimulated to be effective APCs. Donor APCs that are carried passively in the graft express donor alloantigens and may be recognized directly by recipient T cells (direct allorecognition). Additionally, donor alloantigens can be shed by cells in the graft, taken up by the recipient's APCs, and then presented to recipient T cells (indirect allorecognition). The alloantigens on the surface of the APC are recognized by the T-cell receptor (TCR)–CD3 complex on the surface of the T cell. However, optimal T-cell activation occurs only when there is a second or costimulatory signal between the APC and the T cell. Several costimulatory molecules have been identified that function as receptor–ligand pairs on the APC and T-cell surface that mediate adhesion and mutual activation. Among the most well-characterized is CD28 on the T cell, which binds to B7 molecules (CD80, CD86) on the APC. In the absence of this second signal, T cells may become quiescent or even undergo apoptosis. Engagement of the TCR–CD3 complex by APC, followed by costimulatory signals, results in activation of calcineurin in the cytoplasm of the T cell. Calcineurin dephosphorylates an important transcription factor, nuclear factor of activated T cells (NF-AT), allowing it to enter the nucleus and bind to the promoters of interleukin-2 (IL-2) and other cytokines. MAP-kinases are also activated and move to the nucleus to stimulate the promoters of other important cytokines. Secreted IL-2 activates the cell-surface IL-2 receptor (IL-2R), stimulating clonal expansion of T cells. IL-2 (along with other cytokines) produced by these T helper cells stimulates expansion of other cells of the immune system, including other T helper cells, cytotoxic T cells, B cells, and natural killer cells. Engagement of the IL-2R, like many other

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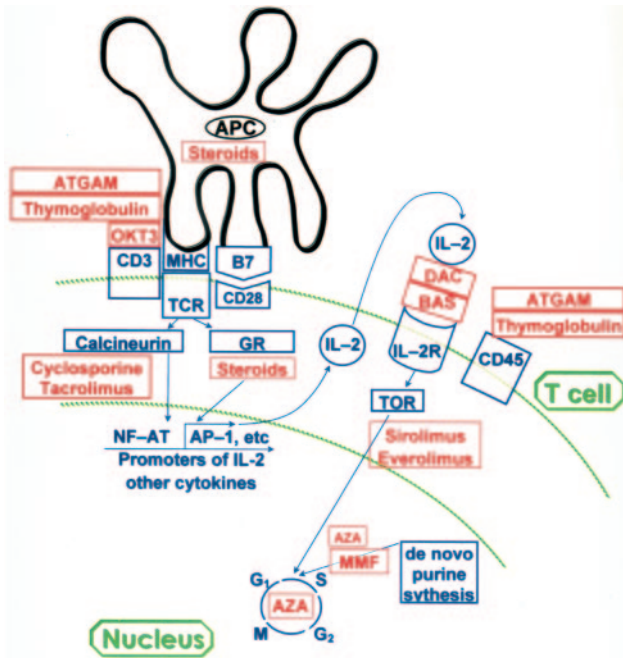
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Immunologic mechanisms leading to graft rejection and sites of action of immunosuppressive drugs. Immunologic mechanisms are shown in blue; immunosuppressive drugs and their site of action are shown in red. Acute rejection begins with recognition of donor antigens that differ from those of recipient by recipient APCs (indirect allorecognition). Donor APCs (carried passively in graft) may also be recognized by recipient T cells (direct allorecognition). Alloantigens carried by APCs are recognized by TCR-CD3 complex on surface of T cell. When accompanied by costimulatory signals between APC and T cells such as B7-CD28, T-cell activation occurs, resulting in activation of calcineurin. Calcineurin dephosphorylates transcription factor NF-AT, allowing it to enter nucleus and bind to promoters of IL-2 and other cytokines. IL-2 activates cell surface receptors (IL-2R), stimulating clonal expansion of T cells (T helper cells). IL-2, along with other cytokines produced by T helper cells, stimulates expansion of other cells of immune system. Activation of IL-2R stimulates TOR, which regulates translation of mRNAs to proteins that regulate cell cycle. Sites of action of individual drugs (highlighted in red) demonstrate multiple sites of action of these drugs, underscoring rationale for combination therapy. AZA indicates azathioprine; MMF, mycophenolate mofetil; GR, glucocorticoid receptor; DAC, daclizumab; and BAS, basiliximab.

growth factor receptors, activates the enzyme target of rapamycin (TOR). TOR regulates the translation of mRNAs to proteins that regulate the cell cycle. The lymphocyte cell cycle requires the de novo synthesis of purines, a process controlled by the enzyme inosine monophosphate dehydrogenase.

Types of Rejection, Timing, and Consequences

Rejection of the transplanted heart is a major cause of morbidity and mortality in the first year after heart transplantation. Rejection is classified as hyperacute, acute cellular, acute humoral (vascular), or chronic. Hyperacute rejection occurs within minutes to hours of the blood flow being reestablished and is caused by preformed antibodies to ABO blood group antigens, HLA, or endothelial antigens. With ABO matching of recipients to donors and prospective cross-matching of patients who have been previously sensi-

tized to HLA, hyperacute rejection is rare. When it does occur, it is catastrophic because preformed antibodies bind to endothelial antigens on the transplanted heart, resulting in activation of complement. An acute inflammatory infiltrate results in fibrinoid necrosis of the vessels of the grafted organ.

Acute cellular rejection may occur at any time after transplantation but is most common in the first 3 to 6 months. It is a T-cell-mediated response with infiltration of lymphocytes and macrophages and resultant myocytolysis. The diagnosis is made by endomyocardial biopsy with a standardized grading scheme ranging from mild to moderate to severe acute rejection.⁹ Moderate rejection by endomyocardial biopsy is associated with mononuclear cell infiltrates and myocytolysis. A diagnosis of moderate rejection generally prompts antirejection therapy that varies according to histological severity (grade of rejection) and hemodynamic function. Patients with acute cellular rejection may have no signs or symptoms but often notice mild symptoms of fatigue or shortness of breath. Signs of right ventricular dysfunction are often noted with elevated jugular venous pressure. More severe rejection may be associated with signs of left heart failure and left ventricular dysfunction. Therapy may include intravenous or oral steroids, monoclonal or polyclonal anti-lymphocyte agents, or an increase or change in oral therapy. The type of therapy generally depends on timing after transplantation, the severity (particularly the severity of hemodynamic compromise), and the protocols of individual centers. In the early 1980s, 70% to 85% of heart transplant recipients experienced acute cellular rejection in the first 6 months after transplantation.¹⁰ More recently, the reported incidence of acute cellular rejection during the first 6 post-operative months is 40% to 70%.^{11,12} Acute cellular rejection does occur after the first 6 months, most often in patients who have had substantial rejection early after transplantation, a recent reduction in immunosuppression, an intercurrent infection, or noncompliance with medication.

Acute humoral (also called vascular) rejection occurs days to weeks after heart transplantation and is initiated by antibodies rather than T cells.^{13,14} The alloantibodies are directed against donor HLA or endothelial cell antigens.¹⁴ Patients at greatest risk of acute humoral rejection include women, patients with a high panel reactive antibody screen and/or a positive cross-match, cytomegalovirus-seropositive recipients, and recipients with sensitization to OKT3.¹⁴ Acute humoral rejection is much less common than acute cellular rejection, occurring in $\approx 7\%$ of patients.¹⁴ Its importance stems from its common association with severe ventricular dysfunction, presumably caused by diffuse ischemia secondary to a lack of coronary vasodilatory reserve. The diagnosis is made by demonstrating immunoglobulin and complement in the vessels of the transplanted heart in an endomyocardial biopsy specimen or by the presence of swollen endothelial cells on hematoxylin and eosin staining.^{13,14} Humoral rejection is treated with intensification of the immunosuppressive regimen but also with therapy directed specifically at either modulating antibody production or removing antibody such as cyclophosphamide immunoglobulin and plasmapheresis. Given the endothelial injury and dysfunction associated with

chronic rejection, it is not surprising that vascular rejection is associated with an increased risk of chronic rejection.¹⁴

Chronic rejection occurs months to years after transplantation.^{15,16} The mechanism is incompletely understood but results from the humoral and cellular consequences of allorecognition. In heart transplant recipients, chronic rejection is also referred to as coronary allograft vasculopathy (CAV) and manifests as diffuse atherosclerosis with myointimal proliferation in the coronary arteries. The diffuse involvement of the coronary arteries results in ischemia and infarction. Angioplasty and coronary bypass surgery are not effective in many patients because of the diffuse nature of the disease.¹⁵ However, angioplasty is frequently performed when focal ischemia is demonstrable. Although the procedure is generally technically successful, the underlying diffuse atherosclerotic process usually progresses rapidly. As many as 50% of heart transplant recipients have angiographically confirmed CAV by 5 years after transplantation, and severe CAV is a major cause of death in patients surviving the first posttransplantation year.^{1,15,16} It remains uncertain whether more intense immunosuppression would ameliorate CAV or whether newer regimens incorporating sirolimus, which may inhibit myointimal proliferation, will prove beneficial.

Immunosuppression

Immunosuppression Regimens

Immunosuppression regimens are generally defined as induction, maintenance, and rejection regimens. Induction therapy is intense perioperative immunosuppressive therapy. Although originally designed to induce tolerance to the graft, this goal has not been realized.⁷ Nonetheless, the concept of induction is useful because it highlights the fact that antidonor responses are typically most vigorous shortly after the transplantation when stimuli such as donor brain death, ischemia/reperfusion, and surgical trauma increase donor antigen expression, thus augmenting the recipient's immune response. The benefits of induction therapy are a marked reduction in rejection in the early postoperative period when graft dysfunction and renal dysfunction are problematic. However, there is increased late rejection after induction therapy is completed. Induction therapy also allows later introduction of calcineurin inhibitors, thus avoiding exacerbation of renal dysfunction. Disadvantages of induction therapy are the increased risk of infection, malignancy, or both and increased cost. Lympholytic agents (ATGAM, Thymoglobulin), generally given for 7 to 14 days postoperatively, have been standard induction drugs. More recently, IL-2R antagonists have been used for induction therapy. In renal and heart transplant recipients, IL-2R antibodies appear to decrease the risk of rejection in the early postoperative period without increasing infection. With the introduction of more potent drugs for maintenance immunosuppression, induction therapy, especially with lympholytic agents, is often reserved for patients at highest risk of rejection or renal failure.

Maintenance therapy generally consists of combination therapy with an antimetabolite, a calcineurin inhibitor, and steroids (Table 1). Maintenance regimens are evolving with

TABLE 1. Common Maintenance Immunosuppressive Regimens

Calcineurin Inhibitor	Antiproliferative Agent	Steroid
Standard regimens		
Cyclosporin or tacrolimus	Mycophenolate mofetil or azathioprine	Prednisone
Newer regimens: TOR inhibitor may replace one standard drug		
Sirolimus or everolimus	Sirolimus or everolimus	

Maintenance immunosuppressive regimens generally consist of a regimen of a calcineurin inhibitor (cyclosporin or tacrolimus) and an antiproliferative agent (mycophenolate mofetil or azathioprine). Prednisone is started in high doses early after transplantation and gradually is tapered to 0 to 5 mg QD by 6 months. Doses of the calcineurin inhibitor also are gradually decreased over time. Newer regimens have substituted a TOR inhibitor for either a calcineurin inhibitor or an antiproliferative agent.

efforts to diminish the nephrotoxicity of calcineurin inhibitors and metabolic toxicity of steroids. Thus, some regimens may add TOR inhibitors to lower doses of calcineurin inhibitors or to eliminate calcineurin inhibitors or steroids. Combination therapy targets several steps in T-cell activation, allowing lower doses of each individual drug (Figure). Specific maintenance regimens vary at individual transplantation centers and are based on age, presensitization, race, and previous rejection because each of these factors determines a patient's risk for rejection. Early maintenance therapy generally consists of a steroid, a calcineurin inhibitor with either cyclosporine (target levels, 300 to 350 ng/mL) or tacrolimus (target levels, 10 to 15 ng/mL), and mycophenolate mofetil at 1 g BID. Most centers have replaced the routine use of azathioprine with mycophenolate mofetil. Therapy is gradually decreased over time, with cyclosporine target levels about 200 ng/mL or tacrolimus target levels at 5 to 10 ng/mL 2 years after transplantation. Because of the long-term side effects, efforts have been made to discontinue maintenance steroid therapy. Prednisone is gradually tapered to 5 mg QD and is discontinued entirely in ≈50% of patients 6 to 12 months after transplantation. Small studies in heart transplant recipients suggest that steroid withdrawal can be accomplished in 30% of patients early (within 6 months of transplantation) and in up to 80% of patients late (24 months) without substantial risk and with an improvement in long-term adverse effects.^{17,18} Thus, heart transplant recipients surviving >1 year are likely to be taking a relatively low dose of a calcineurin inhibitor and mycophenolate mofetil, along with a low dose (5 mg) of prednisone or no steroid at all. Further reductions in immunosuppression are possible in patients who have experienced little rejection. Acute cellular rejection has become less frequent and more easily treated with recent developments in immunosuppressive therapy. However, chronic rejection remains an important problem, as do the long-term side effects caused by these drugs. Recently, it has been suggested that immunosuppressive drugs that prevent acute rejection may also prevent the induction of donor-specific transplantation tolerance.¹⁹ Preliminary data in renal transplant recipients suggest that tolerance may be achievable in a substantial percentage of patients with significantly reduced levels of chronic immunosuppression.²⁰

TABLE 2. Commonly Used Intravenous-Only Immunosuppressive Drugs

Drugs	Trade Name	Pharmacology	Adjustment for Renal/Hepatic Dysfunction	Dosing	Monitoring
Anti-thymocyte globulin*		Elimination by protein degradation and antibody formation to equine (ATGAM) or rabbit (Thymoglobulin) proteins	No		<ul style="list-style-type: none"> • ATGAM requires skin test before first dose. • Premedication† is required. • Monitoring is done by following CD3 counts. • Various targets include^{24,26,30,31} CD3 at 5%–10% baseline, <50 CD3+ cells/mL, 50–100 CD3+ cells/mL. • Repeating daily dose when CD3+ cells increase may decrease number of daily doses, especially with Thymoglobulin.³¹
Polyclonal anti-lymphocyte preparations	ATGAM			10–15 mg · kg ⁻¹ · d ⁻¹ IV over 6–8 h for 3–14 d	
Anti-thymocyte globulin	Thymoglobulin			1.5 mg · kg ⁻¹ · d ⁻¹ IV over 6–8 h for 3–14 d	
Monoclonal* Muromonab CD3	Orthoclone, OKT3	Elimination by protein degradation and binding to target cells with opsonization and phagocytosis	No	5 mg/d for 7–14 d	<ul style="list-style-type: none"> • Premedication† is required.³⁶ • Monitoring CD3+ cells as above • Lower doses may be used if monitoring CD3+ cells.^{26,37} • HAMA may result in an increase of CD3+ cells. • HAMA should be checked before a repeated course is given.
Anti-cytokine receptor antibodies		Elimination via protein degradation similar to IgG ³⁸	No		<ul style="list-style-type: none"> • CD3 counts do not change. • IL-2R + lymphocytes may be measured but are not generally followed clinically. • Rare cases of hypersensitivity to basiliximab have been reported.^{46,47}
Daclizumab	Zenapax			1 mg · kg ⁻¹ · d ⁻¹ within 24 h of transplantation and q 14 d for 4 additional doses. Other dose schedules have been reported. ^{42,45}	
Basiliximab	Simulect			20 mg within 2 h of surgery and 4 d postoperatively	

HAMA indicates anti-human mouse antibodies.

*Generally administered through central line, although Thymoglobulin has been administered through a peripheral line.²⁹

†Premedication for cytokine release syndrome includes antipyretics, intravenous steroids, antihistamines, and H₂ blockers.

Rejection (or rescue) therapy refers to immunosuppressive therapy given to reverse an episode of rejection. The intensity and type of rejection therapy depend on the severity and hemodynamic consequences of the rejection, whether it is thought to be T-cell mediated or humoral, as well as center-specific protocols. Rejection may be treated with an increase in oral therapy, oral or intravenous pulse steroids, a change in oral therapy, or monoclonal or polyclonal anti-lymphocyte agents. Protocols for induction, maintenance, and rejection therapy vary among transplantation centers and often draw on renal transplantation experience, in large part because of the scarcity of randomized, controlled trials in heart transplant recipients.

Immunosuppressive Therapy: General Comments

Immunosuppressive drugs result in 3 categories of outcomes: the desired immunosuppressive effects, the adverse effects of immunodeficiency such as infection and malignancy, and the nonimmune toxicities such as diabetes, hypertension, and renal insufficiency.⁸ Infectious complications, frequent after cardiac transplantation, are a common cause of death in the first year after transplantation and continue to be a significant problem even after the first year.¹ All immunosuppressive drugs contribute to increased risk of infection, with the probable exception of IL-2R antagonists. Malignancy is

another significant problem after cardiac transplantation. Risk factors for malignancy are multifactorial and include impaired immunoregulation, a synergistic effect with other carcinogens such as nicotine or ultraviolet light exposure, and oncogenic viruses such as the Epstein-Barr virus and the papilloma virus.²¹ Lymphoproliferative diseases, skin and lip cancers, and Kaposi's sarcoma have a particularly high incidence relative to the general population. A relatively common cause of death after the first year after transplantation, malignancies account for 24% of deaths after 5 years.¹ All immunosuppressive drugs contribute to the risk of malignancy, with the possible exception of steroids. Data in animals suggest that the antigrowth properties of a new immunosuppressive drug, sirolimus, may result in fewer malignancies.²² The cumulative amount of immunosuppression, especially with OKT3 and polyclonal anti-lymphocyte preparations, is positively correlated with the risk of malignancy.²³ The following discussions of each drug focus on the nonimmune adverse effects.

Immunosuppressive Therapy: Specific Drugs—Intravenous Only

Table 2 summarizes trade names, pharmacology, necessary adjustments for renal or hepatic dysfunction, and dosing and general monitoring guidelines for commonly used intrave-

TABLE 3. Cost of Intravenous Medications Used for Induction Immunosuppression and Rejection⁴⁸

Medication	Dose*	Course of Therapy	Cost per Course,† \$
Muromonab-CD3 (Orthoclone-OKT3)	5 mg	7–14 d	6405–12 810
Antithymocyte immune globulin (ATGAM)	10 mg/kg	7–14 d	6417–12 835
Anti-thymocyte immune globulin (Thymoglobulin)	1.5 mg/kg	7–14 d	10 539–21 078
Daclizumab (Zenapax)	1 mg/kg	Within 24 h before transplantation and then q 14 d for a total of 5 doses	8188
Basiliximab (Simulect)	20 mg	Within 2 h of surgery and then 4 d postoperatively	3238
Methylprednisone (Solumedrol)	250 mg	3 d	14.31
Cytomegalovirus-immune globulin ⁴⁹ (Cytogam)	150 mg/kg	Initially, then at 2, 4, 6, 8 wk	15 382
	100 mg/kg	At 16 and 18 wk	4102
Immune globulin ⁵⁰ intravenous (Polygam)	2 g/kg	1–3 doses	5320–15 960

*Dose based on a 70-kg adult.

†Data are based on average wholesale price in February 2003. Additional administration costs are not included.

nous immunosuppressive drugs. Methyl prednisone, available in both oral and intravenous forms, is included in Part II of this series with corticosteroids.

Table 3 lists the average cost of a typical course of each drug. A table listing common adverse events of both intravenous and oral immunosuppressive drugs is included in Part II.

Anti-Lymphocyte Preparations

There are 2 general types of anti-lymphocyte antibodies, polyclonal and monoclonal.

Polyclonal Anti-Lymphocyte Antibodies

The 2 available formulations of polyclonal anti-lymphocyte antibodies are produced either in horses (ATGAM[®]) or in rabbits (Thymoglobulin[®]).

- **Mechanism of Action.** Polyclonal antibodies (Figure) result in substantial lymphocyte depletion.²⁴ These preparations contain antibodies to many surface T- and B-cell molecules, including HLA.²⁵ Antibodies to CD45, a protein that plays a role in T-cell activation, may be particularly important in reversing rejection and inducing tolerance.²⁶ Treatment results in complement-dependent opsonization and eventual cell lysis and may contribute to apoptosis of these cells. There may be binding to granulocytes and platelets and a reduction of these cells in peripheral blood. The xenogeneic origin of these polyclonal antibodies can induce a host antibody response that results in acute hypersensitivity response or serum sickness on subsequent exposure. Because batches of polyclonal antibodies vary in potency, monitoring of T cells with flow cytometry is helpful in assessing effectiveness and adjusting dosing.
- **Uses and Clinical Trials.** Polyclonal antibodies are used for induction and for the treatment of steroid-resistant rejection in heart transplant recipients. Both ATGAM and Thymoglobulin have Food and Drug Administration (FDA) approval for the management of acute rejection in renal transplant recipients. The few comparisons between ATGAM and Thymoglobulin appear predominantly in the renal transplantation literature. From the limited data available, Thymoglobulin appears to be moderately more efficacious than ATGAM when used for induction therapy or

for treating steroid-resistant rejection.^{27,28} ATGAM is also used in the early perioperative management of patients with worsening renal insufficiency when treatment with calcineurin inhibitors is delayed to prevent the development of acute renal failure.

- **Adverse Effects.** Because both of these agents are foreign proteins, there is a risk of allergic reactions. Urticaria is more common with ATGAM, and fever, chills, and rash may occur with both compounds, especially after the first dose. The cytokine release syndrome, more common with OKT3, can occur with the polyclonal antibodies. Hypertension, diarrhea, and headache are common. Serum sickness can occur, especially with the equine-derived ATGAM. Leukopenia and thrombocytopenia may require either a reduction in dose or termination of therapy. There is an increased incidence of either primary or reactivation cytomegalovirus infections with the use of both monoclonal and polyclonal antibodies, and prophylactic doses of ganciclovir are given during and for up to 3 months after the course of intravenous anti-lymphocyte therapy.²⁶

Monoclonal Anti-Lymphocyte Antibodies

- **Muromonab CD3.** Muromonab-CD3 (OKT3) is a murine antibody that recognizes the epsilon chain of the CD3 molecule on T cells.
- **Mechanism of Action.** CD3 is required for the TCR to generate the intracellular signals that activate T cells.³² The binding of OKT3 to CD3 renders the T cell unable to respond to an antigen challenge or to bind to target cells (Figure). T cells bound to OKT3 are opsonized and removed from the circulation by macrophages in the spleen and liver. Initial binding of OKT3 to the TCR-CD3 complex results in activation of the T cells with release of multiple cytokines. This cytokine release syndrome represents an important aspect in the adverse effect profile of this antibody.
- **Uses and Clinical Trials.** OKT3 has FDA approval for the treatment of acute allograft rejection in renal transplant patients and for the treatment of steroid-resistant acute allograft rejection in cardiac and hepatic transplant patients. In heart transplant recipients, OKT3 has been used primarily for treatment of steroid-refractory rejection, es-

pecially when it is associated with hemodynamic compromise, and for induction therapy in recipients at greater risk of rejection.³³ OKT3 has a 90% rate of complete or partial reversal of biopsy-proven steroid-resistant rejection in heart transplant recipients.³⁴ The experience with induction therapy with OKT3 has not been as promising as the original studies indicated. Although the incidence of early cellular rejection is lower than with conventional triple-drug therapy, there is an increased incidence of late rejection, a higher rate of humoral (vascular) rejection, and no overall benefit at 1 year after cardiac transplantation.³⁵

- **Adverse Effects.** Major adverse reactions with OKT3 are due to either cytokine release or development of antibodies to the mouse immunoglobulin. The cytokine release syndrome is the most dramatic and potentially life-threatening adverse response to this antibody.³⁶ The syndrome usually occurs with the first and second doses of drug, and the incidence greatly diminishes with subsequent doses. Symptoms include fever, chills, rigors, dyspnea, wheezing, chest pain or tightness, headache, nausea, vomiting, and diarrhea. Cardiogenic and noncardiogenic pulmonary edema can occur, and both aseptic meningitis and encephalopathy have been reported. Pulmonary edema is uncommon if fluid overload has been corrected before administration. To prevent major manifestations of this syndrome, antipyretics, intravenous steroids, antihistamines, and occasionally H₂ blockers are routinely prescribed 1 hour before administration of OKT3. As with polyclonal antibodies, routine prophylactic treatment with ganciclovir is recommended.³⁸ Development of anti-mouse antibodies by the recipient prevents a therapeutic benefit and has been associated with an increased incidence of humoral rejection.¹⁴

Anti-Cytokine Receptor Antibodies

Anti-cytokine receptor antibodies used in transplantation are daclizumab and basiliximab. Daclizumab is a humanized anti-IL-2R (CD25) monoclonal antibody that has the murine antigen-binding sequences molecularly engrafted onto a human antibody. Basiliximab is a chimeric (mouse/human) anti-IL-2R monoclonal antibody with mouse variable regions fused to the constant regions of a human IgG.

Mechanism of Action

Both basiliximab and daclizumab bind the α subunit of IL-2R expressed on antigen-activated T cells. This prevents binding of IL-2 to the IL-2R, inhibiting proliferation of T cells.^{40,41} However, this action alone is not sufficient to prevent rejection, and there appear to be other important, although incompletely understood, actions of these antibodies.^{26,42}

Uses and Clinical Trials

Basiliximab and daclizumab are used as induction therapy in many heart transplantation centers. Some centers reserve these agents only for high-risk recipients. Both are FDA approved for the prophylaxis of acute organ rejection in patients receiving renal transplants in a regimen that includes cyclosporine and corticosteroids. One small trial randomized 55 heart transplant recipients receiving prednisone, mycophenolate mofetil, and cyclosporine to daclizumab or no additional therapy.⁴⁰ During the induction period (3 months), acute rejection, defined as an endomyocardial biopsy grade of ≥ 2 , was decreased from 63% to 18% ($P=0.04$). Mortality was not different. The need for anti-lymphocyte therapy and

the frequency of development of anti-HLA antibodies were significantly reduced. Duration of hospitalization, readmission, infections, and malignancy were not different, although there was a trend for the duration of hospitalization to be shorter in the daclizumab group. Several randomized studies in renal transplant recipients have shown similar results, demonstrating a 28% to 37% reduction in biopsy-proven rejection at 6 to 12 months in recipients of a first renal transplant.^{41,43,44} However, a recent, as-yet-unpublished, double-blind, randomized, controlled trial comparing daclizumab with placebo in 434 heart transplant recipients demonstrated an increase in mortality in the daclizumab group.⁴⁵

Adverse Effects

Few serious common adverse events have been reported. Cytokine release syndrome does not occur after administration of these drugs, and there has been no reported increased risk of infection or malignancy.^{40,41,43,44} Hypersensitivity has been reported with initial exposure and reexposure to both basiliximab and daclizumab. The second dose should be withheld if complications such as hypersensitivity occur.^{46–48}

Table 3 describes the costs of intravenous drugs commonly used for induction or antirejection immunosuppression. Methyl prednisolone is included in Table 2 but is described in Part II with other corticosteroids.^{29–31,37,39,49,50}

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References

1. Taylor DO, Edwards LB, Mohacsi PJ, Boucek MM, Trulock EP, Keck BM, Hertz MI. The registry of the International Society for Heart and Lung Transplantation: twentieth official adult heart transplant report—2003. *J Heart Lung Transplant.* 2003;22:616–624.
2. 2000 Annual Report of the U.S. Scientific Registry of Transplant Recipients and the Organ Procurement and Transplantation Network: Transplant Data 1989–1998. Rockville, Md, and Richmond, Va: HHS/HRSA/OSP/DOT and UNOS; 2001.
3. Walden JA, Stevenson LW, Dracup K, Wilmarth J, Kobashigawa J, Moriguchi J. Heart transplantation may not improve quality of life for patients with stable heart failure. *Heart Lung.* 1989;18:497–506.
4. Salyer J, Sneed G, Corley MC. Lifestyle and health status in long-term cardiac transplant recipients. *Heart Lung.* 2001;30:445–457.
5. Turka LA. Normal immune responses. In: Norman DJ, Turka LA, eds. *Primer on Transplantation.* Mr. Laurel, NJ: American Society of Transplantation; 2001:3–15.
6. Krensky A. Immune response to allografts. In: Norman DJ, Turka LA, eds. *Primer on Transplantation.* Mt Laurel, NJ: American Society of Transplantation; 2001:16–25.
7. Denton MD, Magee CC, Sayegh MH. Immunosuppressive strategies in transplantation. *Lancet.* 1999;353:1083–1091.
8. Halloran PF GS. Principles and overview of immunosuppression. In: Norman DJ, Turka LA, eds. *Primer on Transplantation.* Mt Laurel, NJ: American Society of Transplantation; 2001:87–98.
9. Winters GL, Marboe CC, Billingham ME. The International Society for Heart and Lung Transplantation grading system for heart transplant biopsy specimens: clarification and commentary. *J Heart Lung Transplant.* 1998; 17:754–760.
10. Hunt S. Complications of heart transplantation. *J Heart Transplant.* 1983;3: 70–78.
11. Eisen HJ, Hobbs RE, Davis SF, Laufer G, Mancini DM, Renlund DG, Valentine H, Ventura H, Vachieri JL, Bourge RC, Canver CC, Carrier M, Costanzo MR, Copeland J, Dureau G, Frazier OH, Dorent R, Hauptman PJ, Kells C, Master R, Michaud JL, Paradis I, Smith A, Vanhaecke J, Mueller EA, et al. Safety, tolerability and efficacy of cyclosporine microemulsion in heart transplant recipients: a randomized, multicenter, double-blind com-

- parison with the oil based formulation of cyclosporine: results at six months after transplantation. *Transplantation*. 1999;68:663–671.
12. Kobashigawa J, Miller L, Renlund D, Mentzer R, Alderman E, Bourge R, Costanzo M, Eisen H, Dureau G, Ratkovec R, Hummel M, Ipe D, Johnson J, Keogh A, Mamelok R, Mancini D, Smart F, Valentine H. A randomized active-controlled trial of mycophenolate mofetil in heart transplant recipients: Mycophenolate Mofetil Investigators. *Transplantation*. 1998;66:507–515.
 13. Hammond EH, Yowell RL, Nunoda S, Menlove RL, Renlund DG, Bristow MR, Gay WA Jr, Jones KW, O'Connell JB. Vascular (humoral) rejection in heart transplantation: pathologic observations and clinical implications. *J Heart Transplant*. 1989;8:430–443.
 14. Michaels PJ, Espejo ML, Kobashigawa J, Alejos JC, Burch C, Takemoto S, Reed EF, Fishbein MC. Humoral rejection in cardiac transplantation: risk factors, hemodynamic consequences and relationship to transplant coronary artery disease. *J Heart Lung Transplant*. 2003;22:58–69.
 15. Behrendt D, Ganz P, Fang JC. Cardiac allograft vasculopathy. *Curr Opin Cardiol*. 2000;15:422–429.
 16. Young JB. Perspectives on cardiac allograft vasculopathy. *Curr Atheroscler Rep*. 2000;2:259–271.
 17. Taylor DO, Bristow MR, O'Connell JB, Price GD, Hammond EH, Doty DB, Karwande SV, Gay WA Jr, Jones KW, Lappe D, Renlund DG. Improved long-term survival after heart transplantation predicted by successful early withdrawal from maintenance corticosteroid therapy. *J Heart Lung Transplant*. 1996;15:1039–1046.
 18. Kobashigawa JA, Stevenson LW, Brownfield ED, Gleeson MP, Moriguchi JD, Kawata N, Minkley R, Drinkwater DC, Laks H. Corticosteroid weaning late after heart transplantation: relation to HLA-DR mismatching and long-term metabolic benefits. *J Heart Lung Transplant*. 1995;14:963–967.
 19. Smiley ST, Csizmadia V, Gao W, Turka LA, Hancock WW. Differential effects of cyclosporine A, methylprednisolone, mycophenolate, and rapamycin on CD154 induction and requirement for NFkappaB: implications for tolerance induction. *Transplantation*. 2000;70:415–419.
 20. Starzl TE, Murase N, Abu-Elmagd K, Gray EA, Shapiro R, Eghtesad B, Corry RJ, Jordan ML, Fontes P, Gayowski T, Bond G, Scantlebury VP, Potdar S, Randhawa P, Wu T, Zeevi A, Nalesnik MA, Woodward J, Marcos A, Trucco M, Demetris AJ, Fung JJ. Tolerogenic immunosuppression for organ transplantation. *Lancet*. 2003;361:1502–1510.
 21. Penn I. Post-transplant malignancy: the role of immunosuppression. *Drug Saf*. 2000;23:101–113.
 22. Guba M, von Breitenbuch P, Steinbauer M, Koehl G, Flegel S, Hornung M, Bruns CJ, Zuelke C, Farkas S, Anthuber M, Jauch KW, Geissler EK. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. *Nat Med*. 2002;8:128–135.
 23. Swinnen LJ, Costanzo-Nordin MR, Fisher SG, O'Sullivan EJ, Johnson MR, Heroux AL, Dizikes GJ, Pifarre R, Fisher RI. Increased incidence of lymphoproliferative disorder after immunosuppression with the monoclonal antibody OKT3 in cardiac-transplant recipients. *N Engl J Med*. 1990;323:1723–1728.
 24. Bonnefoy-Berard N, Revillard JP. Mechanisms of immunosuppression induced by antithymocyte globulins and OKT3. *J Heart Lung Transplant*. 1996;15:435–442.
 25. Bourdage JS, Hamlin DM. Comparative polyclonal antithymocyte globulin and antilymphocyte/antilymphoblast globulin anti-CD antigen analysis by flow cytometry. *Transplantation*. 1995;59:1194–1200.
 26. George J. In: Kirklin JKYJ, McGiffen DC, eds. *Immunosuppressive Modalities in Heart Transplantation*. New York, NY: Churchill Livingstone; 2002:390–463.
 27. Gaber AO, First MR, Tesi RJ, Gaston RS, Mendez R, Mulloy LL, Light JA, Gaber LW, Squiers E, Taylor RJ, Neylan JF, Steiner RW, Knechtle S, Norman DJ, Shihab F, Basadonna G, Brennan DC, Hodge EE, Kahan BD, Kahan L, Steinberg S, Woodle ES, Chan L, Ham JM, Schroeder TJ, et al. Results of the double-blind, randomized, multicenter, phase III clinical trial of Thymoglobulin versus ATGAM in the treatment of acute graft rejection episodes after renal transplantation. *Transplantation*. 1998;66:29–37.
 28. Brennan DC, Flavin K, Lowell JA, Howard TK, Shenoy S, Burgess S, Dolan S, Kano JM, Mahon M, Schmitzler MA, Woodward R, Irish W, Singer GG. A randomized, double-blinded comparison of Thymoglobulin versus ATGAM for induction immunosuppressive therapy in adult renal transplant recipients. *Transplantation*. 1999;67:1011–1018.
 29. Wiland AM, Fink JC, Philopophe B, Farney AC, Schweitzer EJ, Colonna JO, Weir MR, Bartlett ST. Peripheral administration of thymoglobulin for induction therapy in pancreas transplantation. *Transplant Proc*. 2001;33:1910.
 30. Abouna GM, al-Abdullah IH, Kelly-Sullivan D, Kumar MS, Loose J, Phillips K, Yost S, Seirka D. Randomized clinical trial of antithymocyte globulin induction in renal transplantation comparing a fixed daily dose with dose adjustment according to T cell monitoring. *Transplantation*. 1995;59:1564–1568.
 31. Krasinskas AM, Kreisel D, Acker MA, Bavaria JE, Pochettino A, Kotloff RM, Arcasoy S, Blumenthal N, Kamoun M, Moore JS, Rosengard BR. CD3 monitoring of antithymocyte globulin therapy in thoracic organ transplantation. *Transplantation*. 2002;73:1339–1341.
 32. Caillat-Zucman S, Blumenfeld N, Legendre C, Noel LH, Bach JF, Kreis H, Chatenoud L. The OKT3 immunosuppressive effect: in situ antigenic modulation of human graft-infiltrating T cells. *Transplantation*. 1990;49:156–160.
 33. Hooks MA, Wade CS, Millikan WJ Jr. Muromonab CD-3: a review of its pharmacology, pharmacokinetics, and clinical use in transplantation. *Pharmacotherapy*. 1991;11:26–37.
 34. Haverty TP, Sanders M, Sheahan M. OKT3 treatment of cardiac allograft rejection. *J Heart Lung Transplant*. 1993;12:591–598.
 35. Adamson R, Bispo E, Dychter S, Dembitsky W, Moreno-Cabral R, Jaski B, Gordon J, Hoagland P, Moore K, King J, Andrews J, Rich M, Daily PO. Long-term outcome with the use of OKT3 induction therapy in heart transplant patients: a single-center experience. *Transplant Proc*. 1998;30:1107–1109.
 36. Chatenoud L, Ferran C, Reuter A, Legendre C, Gevaert Y, Kreis H, Franchimont P, Bach JF. Systemic reaction to the anti-T-cell monoclonal antibody OKT3 in relation to serum levels of tumor necrosis factor and interferon-gamma [corrected]. *N Engl J Med*. 1989;320:1420–1421.
 37. Midtvedt K, Fauchald P, Lien B, Hartmann A, Albrechtsen D, Bjerkely BL, Leivestad T, Brekke IB. Individualized T cell monitored administration of ATG versus OKT3 in steroid-resistant kidney graft rejection. *Clin Transplant*. 2003;17:69–74.
 38. Merigan TC, Renlund DG, Keay S, Bristow MR, Starnes V, O'Connell JB, Resta S, Dunn D, Gamberg P, Ratkovec RM, et al. A controlled trial of ganciclovir to prevent cytomegalovirus disease after heart transplantation. *N Engl J Med*. 1992;326:1182–6.
 39. Loertscher R. The utility of monoclonal antibody therapy in renal transplantation. *Transplant Proc*. 2002;34:797–800.
 40. Beniaminovitz A, Itescu S, Lietz K, Donovan M, Burke EM, Groff BD, Edwards N, Mancini DM. Prevention of rejection in cardiac transplantation by blockade of the interleukin-2 receptor with a monoclonal antibody. *N Engl J Med*. 2000;342:613–619.
 41. Kahan BD, Rajagopalan PR, Hall M. Reduction of the occurrence of acute cellular rejection among renal allograft recipients treated with basiliximab, a chimeric anti-interleukin-2-receptor monoclonal antibody: United States Simulect Renal Study Group. *Transplantation*. 1999;67:276–284.
 42. ter Meulen CG, Baan CC, Hene RJ, Hilbrands LB, Hoitsma AJ. Two doses of daclizumab are sufficient for prolonged interleukin-2Ralpha chain blockade. *Transplantation*. 2001;72:1709–1710.
 43. Vincenti F, Kirkman R, Light S, Bumgardner G, Pescovitz M, Halloran P, Neylan J, Wilkinson A, Ekberg H, Gaston R, Backman L, Burdick J. Interleukin-2-receptor blockade with daclizumab to prevent acute rejection in renal transplantation: Daclizumab Triple Therapy Study Group. *N Engl J Med*. 1998;338:161–165.
 44. Nashan B, Moore R, Amlot P, Schmidt AG, Abeywickrama K, Soullou JP. Randomised trial of basiliximab versus placebo for control of acute cellular rejection in renal allograft recipients: CHIB 201 International Study Group. *Lancet*. 1997;350:1193–1198.
 45. Important Drug Warnings. Basel, Switzerland: Hoffman-La Roche, Inc; 2003.
 46. Leonard PA, Woodside KJ, Gugliuzza KK, Sur S, Daller JA. Safe administration of a humanized murine antibody after anaphylaxis to a chimeric murine antibody. *Transplantation*. 2002;74:1697–1700.
 47. Basiliximab [package insert]. East Hanover, NJ: Novartis; 2003.
 48. Cardinal Health. Cardinal Health Distribution Database. Available at: <http://www.cardinal.com>. Accessed November 19, 2004.
 49. Cytomegalovirus immune globulin intravenous [CytoGam package insert]. Gaithersburg, Md: MedImmune, Inc; 2000.
 50. Jordan SC, Quartel AW, Czer LS, Admon D, Chen G, Fishbein MC, Schwieger J, Steiner RW, Davis C, Tyran DB. Posttransplant therapy using high-dose human immunoglobulin (intravenous gammaglobulin) to control acute humoral rejection in renal and cardiac allograft recipients and potential mechanism of action. *Transplantation*. 1998;66:800–805.